

The Search for Infectious Causes of Human Cancers: Where and Why (Nobel Lecture)**

Harald zur Hausen*

cancer research · Nobel Lecture · papillomaviruses · virology

Slightly more than 20 % of the global cancer burden can currently be linked to infectious agents, including viruses, bacteria, and parasites. In this Review the reasons for their relatively late discovery are analyzed, and epidemiological observations that may point to an involvement of additional infectious agents in specific human cancers are highlighted. Emphasis is placed on hematopoietic malignancies, breast and colorectal cancers, as well as basal cell carcinomas of the skin and lung cancers in nonsmokers.

1. Introduction

1.1. Present State of the Global Cancer Burden

Currently a larger number of infectious agents have been identified which either cause or contribute to specific human cancers.^[1a] They include two members of the herpes virus family (Epstein-Barr virus and human herpesvirus type 8), high-risk and low-risk human papillomaviruses (HPV), hepatitis B and C viruses, a recently identified human polyomavirus, Merkel cell polyomavirus,^[2] the human T-lymphotropic retrovirus type 1 (HTLV-1), and human immunodeficiency viruses (HIV) types 1 and 2. In addition, human endogenous retroviruses have been suspected to play a role in human cancers. Besides viruses, other pathogens have also been

identified. They include the bacterium *Helicobacter pylori*, a major contributor to gastric cancer, and parasitic infections, here in particular *Schistosoma hematobium*, a major cause of bladder cancer in Egypt, and liver flukes. The latter, *Opisthorchis viverrini* and *Clonorchis sinensis*, are important factors for cholangiocarcinomas and hepatocellular carcinomas in South-Eastern Thailand and Southern China. Figure 1 shows an estimate of the present contribution of infectious agents to the global cancer incidence.

It is important to note that there exist vast gender differences in the global role of papillomaviruses in human cancers. This is mainly due to the role of this virus family in the induction of cancer of the cervix. More than 50 % of cancers linked to infections in females are caused by HPV infections. In males, only approximately 4.3 % of cancers have been linked to this virus family.

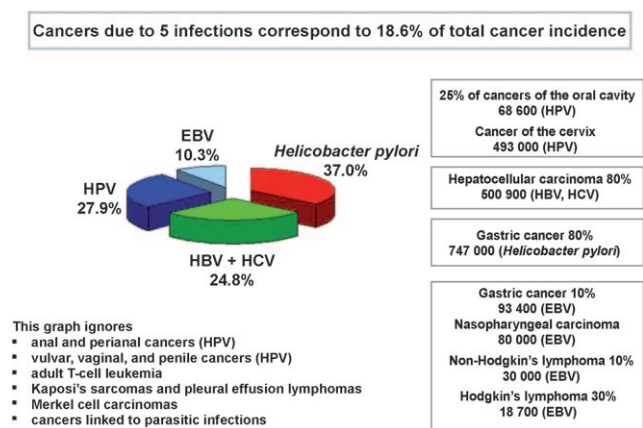


Figure 1. Estimated annual global cancer incidence caused by infections.^[1]

2. Problems in Identifying Infectious Agents Involved in Human Cancer Induction

2.1. Why Has it Been so Difficult to Identify Infectious Agents as Causative Factors for Human Tumors?

The search for an infectious cause of at least some human cancers dates back to the second half of the nineteenth century.^[1a] Yet, the first hints for a role of infectious agents in

[*] Prof. H. zur Hausen
Deutsches Krebsforschungszentrum
Im Neuenheimer Feld 280, 60120 Heidelberg (Germany)
E-mail: zurhausen@dkfz-heidelberg.de

[**] Copyright© The Nobel Foundation 2008. We thank the Nobel Foundation, Stockholm, for permission to print this lecture.

human cancers date back to the beginning of the 20th century, when *Schistosoma* infections in Egypt and liver flukes in Eastern Europe and Asia were suspected to play a role in the development of bladder and liver cancers. Despite intensive search, it took approximately 65 additional years before further evidence was obtained, namely by linking a specific virus, the Epstein–Barr virus, to two human cancers, Burkitt's lymphoma and nasopharyngeal carcinoma. During the past three or four decades progress has been more rapid, linking currently about 20% of the global cancer incidence to infectious events.

Why has it been so difficult to identify infectious agents as causative factors for human cancers? Several reasons seem to provide an explanation:

1. Because no human cancer arises as the acute consequence of infection. The latency periods between primary infection and cancer development are frequently in the range of 15 to 40 years. The X-chromosome-linked lymphoproliferation (XLLP) represents a rare exception. Based on a specific host-cell mutation, the Epstein–Barr virus here causes an acute lymphoproliferative disease.
2. Besides some rare exceptions, no synthesis of the infectious agents occurs in cancer cells.
3. Most of the infections linked to human cancers are common in human populations—they are ubiquitous. They were present during the whole human evolution. Yet, only a small proportion of infected individuals develops the respective cancer type.
4. Mutations in host-cell genes or within the viral genome are mandatory for malignant conversion.
5. Chemical (for example, aflatoxin) and physical carcinogens (for example, ultraviolet light in Epidermodysplasia verruciformis) act usually as mutagens. They facilitate the selection of specific mutations and frequently act synergistically with carcinogenic infectious agents.
6. Some infectious agents act as indirect carcinogens, without persistence of their genes within the respective cancer cells (HIV, *Helicobacter pylori*, *Schistosoma hematobium*, hepatitis C and B).

Among all these factors, the ubiquity of most of these infections and the long time periods required for malignant transformation were the main reasons for the remarkable difficulties in identifying their carcinogenic functions.



Harald zur Hausen, born March 11, 1936 in Gelsenkirchen (Germany), studied Medicine in Bonn, Hamburg, and Düsseldorf, where in 1960 he completed his PhD. In 1969 he completed his Habilitation at the University of Würzburg, before becoming Professor for Clinical Virology at the University of Erlangen–Nürnberg in 1972. In 1977 he moved to the University of Freiburg. From 1983 to 2003 he was president of the DKFZ in Heidelberg.

2.2. Epidemiology Provided Hints for a Successful Search

2.2.1. Geographic Coincidence

Geographic coincidence of a specific infection (hepatitis B) and of liver cancer led to the original suspicion that this infection may predispose to the subsequent development of hepatocellular carcinomas.^[1a] The additional contribution of a chemical carcinogen was also suspected based on similar observations. Figure 2 reveals the geographic distribution of

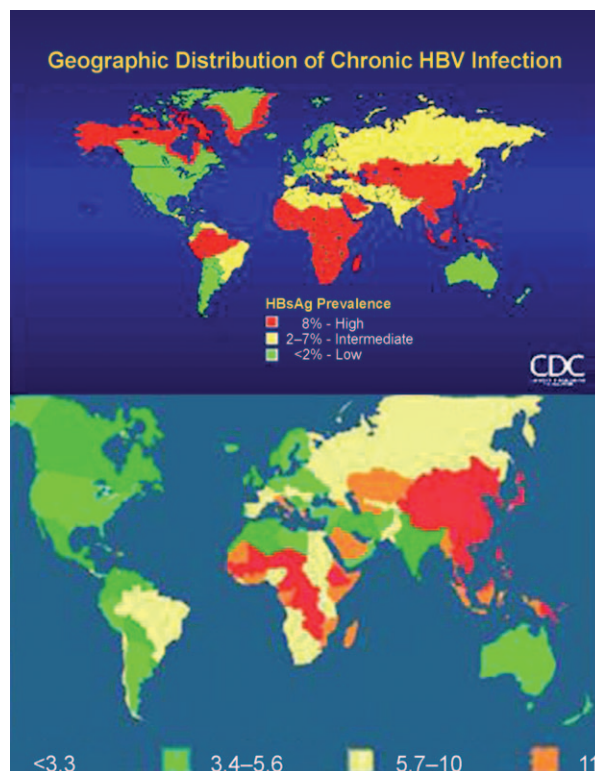


Figure 2. Geographic distribution of hepatitis B infections (top) and of hepatocellular carcinomas (bottom). Modified from figures provided by CDC and Globocan 2002.

hepatitis B virus infections and hepatocellular carcinomas. Geographic clustering of specific cancers may, however, also result from other causes: Countries with a high rate of heavy smokers also experience a high incidence of lung cancer. The intensive solar exposure of Caucasian populations in Australia, South Africa, and South America is responsible for a high percentage of skin cancer patients.

2.2.2. Regional Clustering of Cases

Regional clustering of specific cancer types triggered some investigations on a potential role of infectious agents in these malignant proliferations. Burkitt's lymphoma in equatorial Africa represents one of the most illustrative examples. Burkitt noted the apparent dependence of tumor incidence on climatic conditions and altitude, and described the regional correlation with holoendemic *Plasmodium falciparum* infec-

tions.^[3] As a consequence, he speculated that the tumor might be due to a viral infection, transmitted by an arthropod vector, possibly the same carrying malaria parasites.

Nasopharyngeal carcinoma, occurring at high frequency in specific regions of South-East Asia, represents another example. Adult T-cell leukemia in the coastal regions of Southern Japan, cholangiocarcinomas in South-East Thailand, and bladder cancer in the Nile Delta or along the Nile river also raised early suspicions for an infectious origin. These observations resulted in speculations, but they could not prove the underlying hypothesis by themselves.

2.2.3. Dependence on Sexual Contacts

If one disregards the occurrence of scrotum cancer in chimney sweepers, the early studies of Rigoni-Stern in Verona, Italy, pointing in 1842 to a role of sexual contacts in the causation of cervical cancer, represent a particularly interesting example of suspected contact transmission of a human cancer. It took another 140 years before the viral infections were identified that caused this frequent cancer in women. These observations led to the identification of additional anogenital and oral cancers linked to the same virus infections.

2.2.4. Cancers Arising under Immunosuppression

Epidemiological surveys identified immunosuppression as a condition resulting in the appearance of remarkably specific forms of cancer. Many of those malignancies have now been shown to be caused by reactivated viruses, whose oncogenic potential is usually suppressed by immunological reactions. The most prominent tumors arising here are Epstein-Barr virus caused B-cell lymphomas, Kaposi's sarcomas linked to human herpesvirus type 8 reactivation, and Merkel cell carcinomas of the skin associated with a novel polyomavirus. The initial discovery of the viral origin of cervical cancer and its precursor lesions was not based on the moderately enhanced incidence under immunosuppression. Specific types of common warts also occur as a nonmalignant proliferative condition at high frequency in immunosuppressed patients, mainly containing genus-Beta papillomaviruses. The viral origin of basal and squamous cell carcinomas of the skin, frequently found in these patients, remains up to now controversial.

3. Mechanistic Aspects of Cancer Induction by Infections

Figure 3 lists identified mechanisms by which infections may contribute to cancer development. The expression of specific viral oncogenes as a mandatory precondition for the maintenance of the malignant phenotype has been identified as a direct contribution to human carcinogenesis.^[1a] A novel mode of direct viral carcinogenesis has probably been identified in Merkel cell carcinomas, where functional inactivation of the helicase part of the large T-antigen of the Merkel cell polyomavirus renders the viral DNA replication-

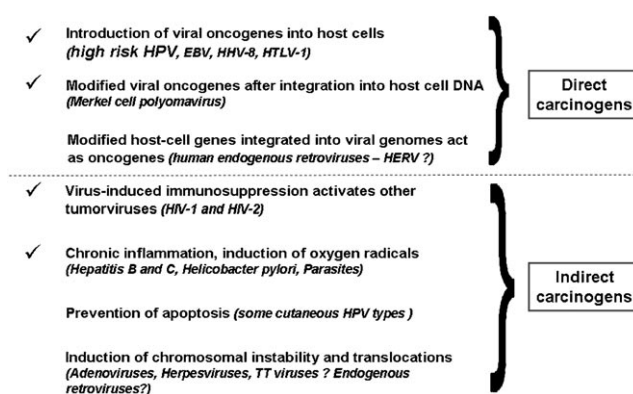


Figure 3. Summary of identified mechanisms by which infections either directly or indirectly contribute to carcinogenesis. Mechanistic contributions of infections to human cancers have been marked.

incompetent.^[4] Viral DNA persisting in normal tissues seems to retain replication competence.

The most prominent indirect infectious carcinogens are agents causing immunosuppression or inducing, by inflammatory reactions, reactive oxygen species. Whereas the mechanism of immunosuppression induced by human immunodeficiency viruses (HIV) or after organ transplantation is reasonably well understood, the accurate mechanism by which hepatitis B and C viruses, *helicobacter pylori*, and carcinogenic parasitic infections contribute to cancer still remains somewhat obscure.

4. Where Is it Worthwhile To Search for an Infectious Etiology of Human Cancers not yet Linked to Infections?

When we summarize infectious agents that have been discovered during the past 15 years, it is interesting to note that several novel viruses belonging to potentially carcinogenic virus families have even been identified during the past 2 years (Figure 4). This raises the suspicion that additional links to novel or already identified infectious agents to cancers will become apparent, hitherto not linked to infections. Thus, it appears worthwhile to search for cancer-related epidemiologic observations that may point to the involvement of infectious agents in cancers hitherto not linked to infections. The following section will summarize some hypotheses and considerations based on these reports.

4.1. Cancers Occurring under Immunosuppression

A review published in 2006 by Vajdic and colleagues^[5] demonstrates a larger number of cancers occurring at increased frequency under immunosuppression after kidney transplantation. Kaposi's sarcoma, mainly found in HIV-infected patients, stands out and is found about 200-fold increased in these patients in comparison to non-infected controls (Figure 5). The most interesting part of Figure 5

Year	Virus	Symptoms	Natural Host
1994	Sabia virus	Hemorrhagic fever	Rodents
1994	Hum. Herpesvirus 8	Kaposi's sarcoma	Humans
1994	Hendraviruses	Encephalitis	Bats, horses
1997	Influenza H5N1	Avian flue	Birds
1997	TT viruses	?	Humans
1998	Nipah virus	Encephalitis	Bats, pigs
2003	SARS Coronavirus	SARS	Chinese bushcat
2005	Bocavirus (parvovirus)	Acute wheezing	Humans
2005	New coronavirus	Respiratory symptoms	Humans
2007	KI-polyomavirus	?	Humans
2007	WU-polyomavirus	?	Humans
2008	MC-polyomavirus	Merkel-tumor	Humans
2008	Lymphotrop. polyomavirus	Periph. blood PML patients	Humans

Within the same time period at least 30 novel types of human papillomaviruses have been identified

Figure 4. "New" human pathogenic viruses (1994–2008). The light arrows identify important human pathogens or a whole novel virus family (TT viruses). The dark arrows point to established or potentially oncogenic virus isolates.

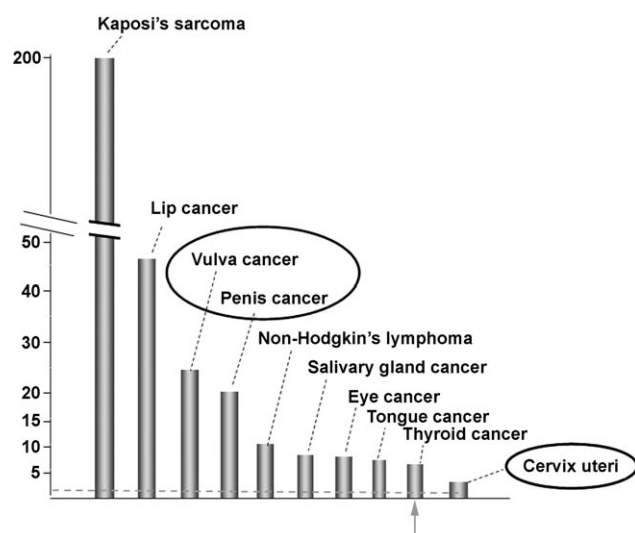


Figure 5. Some of the most frequently occurring cancers occurring after kidney transplantation.^[5] The dotted line indicates the incidence in immunocompetent patients.

appears to be the seven- to eightfold higher rate of vulva and penile cancer in comparison to cancer of the cervix. The vast majority of cervical cancers are caused by high-risk human papillomavirus (HPV) infections. In vulva and penile cancers only 30–50 % seems to be linked to the same HPV infections. The etiology of 50–70 % of these cancers is unknown. Interestingly, the age distribution of HPV-positive and HPV-negative vulva and penile cancers differs in that the negative tumors regularly occur in older age groups. Thus, the negative group require attention as possible candidates for an unknown viral etiology. Unidentified types of HPVs or novel polyomaviruses may represent interesting candidates. Salivary gland, eye, thyroid, and tongue cancers also deserve attention.

4.2. Cancers not Elevated or even Reduced after Immunosuppression

4.2.1. Breast Cancer as an Example

Some cancers do not show an increased incidence during immunosuppression. Indeed, immunosuppression may even possess a protective effect for some of these tumors. Those cancers are shown in Figure 6. Besides prostate, rectum, and

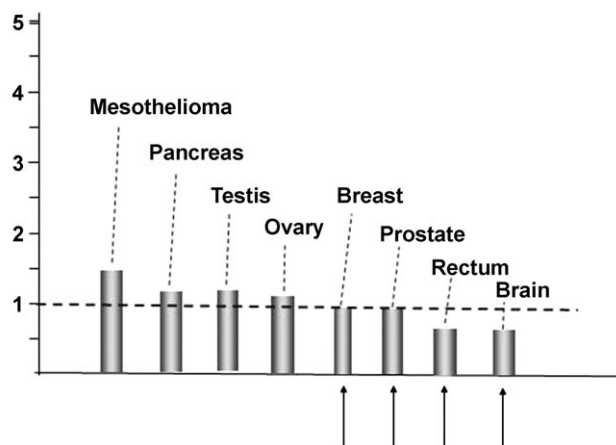


Figure 6. Cancer incidence marginally or not affected during immunosuppression after kidney transplantation.^[5]

brain tumors, human breast cancer represents a particularly intriguing malignancy, because murine mammary cancer is also not increased under immunosuppression. This latter tumor is caused by a retrovirus infection, the murine mammary tumor virus (MMTV).

In murine mammary tumors, the mechanism of a slightly protective effect exerted by immunosuppression is partially understood.^[1a] It is outlined schematically in Figure 7. The primary infection occurs via the milk of the infected mother. The virus reaches the Peyer's patches where it infects B and T lymphocytes. Superantigen induction in the infected cells

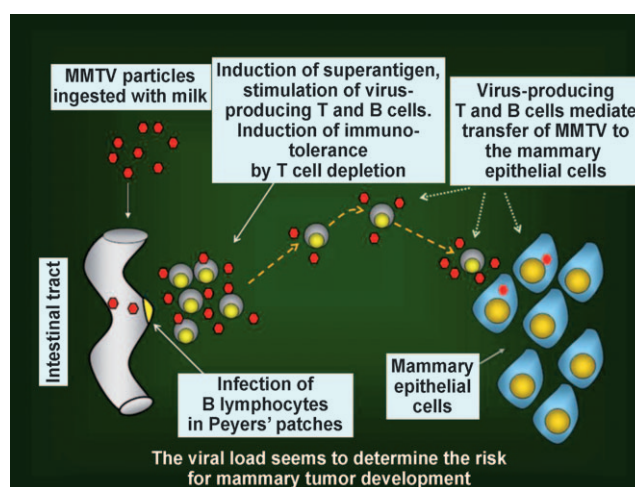


Figure 7. Schematic outline of events following infection of newborn mice with murine mammary tumor virus.^[1a]

leads to reactive T-cell depletion and immunotolerance. The superantigen-expressing cells produce high quantities of infectious MMTV; this substantially increases the risk for the infection of mammary tissue. Specific integration of the MMTV proviral DNA in the mammary cells emerges as the prime risk factor for the resulting mammary carcinomas. Immunosuppression of such infected animals apparently interferes with the emergence of superantigen-producing T and B lymphocytes and, as a consequence, suppresses virus production, which in turn decreases the risk of cancer development.

Is it possible that a similar mechanism contributes to human mammary cancer? A few data seem to support this notion. They may point to a possible involvement of a specific subgroup of human endogenous retroviruses (HERV) in this malignancy. At least 8% of our genome consists of retroviral sequences acquired in the course of human evolution. Although the vast majority of these sequences do no longer reveal functional open reading frames, members of one subgroup, HERV-K, which entered our germline approximately 800 000 years ago, are still able to code for complete, although non-infectious virus particles. Retroviral gag and env transcripts of the 22q11.21 region are found in these particles.^[6] Correction of stop codons in HERV-K sequences resulted even in the reconstitution of infectious HERV-K viruses.^[7,8] HERV-K expression also becomes activated by other virus infections: HIV infections activate HERV-K sequences.^[9] Similarly Epstein–Barr virus infections result in the induction of HERV-K superantigen.^[10–12] Epstein–Barr virus containing Burkitt's lymphoma cells occasionally reveal particles strongly resembling retroviral type A structures upon induction by the tumor-promoting phorbol ester TPA.^[13] Typical structures are shown in Figure 8.

Some recent reports may further stress a potential role of reactivated HERV-K viruses in the pathogenesis of human breast cancer: an antigen-specific immune response was demonstrated in breast-cancer patients.^[14] In addition, breast-cancer patients, HIV-associated lymphomas, non-

HIV-associated lymphomas, and HIV-associated Hodgkin's lymphomas reveal about sevenfold elevated concentrations of HERV-K (HML-2) RNA in their plasma when compared to healthy controls.^[15] The RNA titers in lymphoma patients in remission returned to control values.

Although the available data seem to support a potential role of endogenous retroviruses in human breast cancer, they certainly do not prove it. Other agents may also contribute to at least a proportion of these cancers. A possible link of red meat consumption in relation to breast cancer and a potential involvement of other viral factors will be discussed in connection with a subsequent topic. Nevertheless, human breast cancer remains an interesting candidate for a viral etiology.

4.3. Cancer Incidence Influenced by Infections

The risk for some cancers seems to be influenced by other infections which neither directly contribute to carcinogenesis nor induce long-lasting immunosuppression.

4.3.1. Basal Cell Carcinomas in Pox Scars

Figure 9 reveals an example of multiple basal cell carcinomas arising 20 years in a smallpox vaccination scar. This does not represent a solitary observation, since a larger number of basal cell carcinomas and also melanomas, squamous cell carcinomas, and a few more rarely occurring malignancies (dermatofibrosarcoma protuberans, fibrosarcoma, and malignant fibrous histiocytomas) have also been reported to occur in smallpox vaccination scars.^[16] They are summarized in Figure 10.

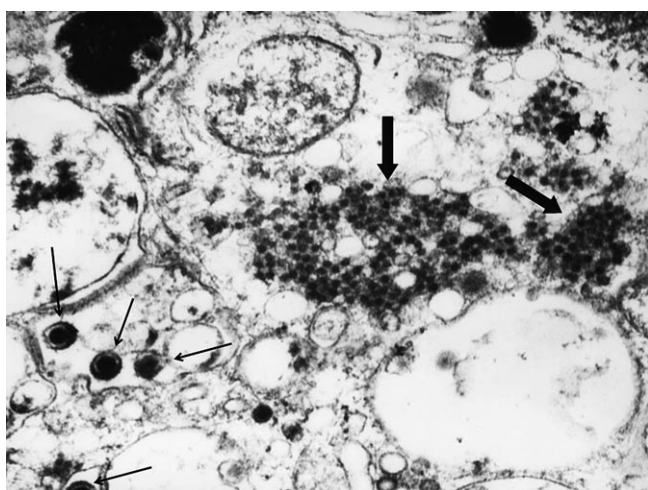


Figure 8. Epstein–Barr virus particles (thin arrows) and two clusters of A-type particle-like structures (thick arrows) in a TPA-treated Burkitt's lymphoma cell.

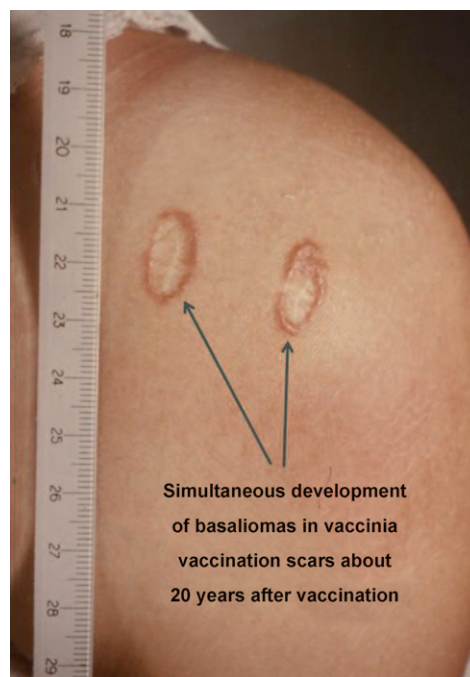


Figure 9. Multifocal basal cell carcinomas in pox scars.

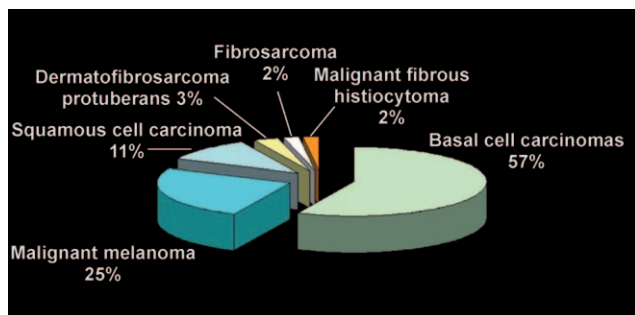


Figure 10. Malignant tumors arising in vaccinia virus vaccination scars.^[16]

Prior to the eradication of smallpox infections, vaccines against these infections were prepared by inoculating vaccinia virus into the scarified skin of calves and harvesting the skin crusts containing the vaccinia virus particles. It is possible that these preparations contained contaminating bovine viruses. Previously it has been demonstrated that vaccinia virus infections cause amplification of persisting polyoma type virus genomes.^[17] This may increase the likelihood for contaminations with bovine members of this virus family. Persisting papillomavirus DNA would be also affected in cells replicating vaccinia virus.^[18]

The published data permit several interpretations:

- Vaccinia virus infection of calf skin resulted in the activation of specific cattle viruses whose subsequent inoculation into humans as contamination represented a risk factor for subsequent local cancer development;
- Vaccinia virus infection of the human skin resulted in local activation of potentially oncogenic human viruses, increasing the risk for cancer development 20–60 years later;
- Early inflammatory reactions induced by this vaccination resulted in mutational events resulting in some cases in the simultaneous appearance of multifocal cancers.

Although other interpretations still remain possible, and basal cell carcinomas have also occasionally been observed in other nonvaccination scars, the observations described here should promote studies on a possible viral role in the initiation of these malignant proliferations

4.3.2. Hematopoietic Malignancies

As shown in Figure 11, a number of human viruses turn out to be oncogenic when inoculated into newborn rodents. Intracerebral infections by JC virus are able to induce astrocytomas in adult owl monkeys.^[19] For obvious reasons, the reverse question, whether specific animal viruses are also able to induce tumors in humans, has not yet been carefully investigated.^[20] Yet, we are living in close contact with domestic animals and regularly handle their products. This is particularly interesting because contact with cattle and consumption of red meat have been identified as risk factors for specific human malignancies. Contact with cattle has also frequently been considered as a risk factor for hematopoietic malignancies, in particular childhood acute lymphocytic leukemias.^[21]

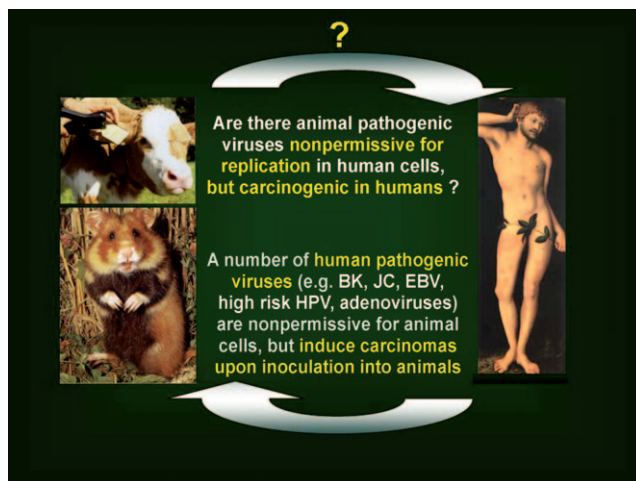


Figure 11. Some human viruses are carcinogenic for several animal species. Do animal viruses exist that are potentially carcinogenic in humans?

4.3.2.1. Risk and Protective Factors

In the following, the reasons for considering childhood leukemias as potential candidates for an infectious etiology will be briefly summarized. A more detailed account has been published recently.^[21] Some protective factors as well as several risk factors for this malignancy are presented in Figure 12.

Repeatedly reported protective factors for childhood leukemias:	Risk factors for childhood leukemias
<div>Multiple infections in early childhood</div> <div>Underprivileged social status</div> <div>Crowded household, many siblings</div> <div>Inverse risk with birth order</div> <div>More than 6 months of breastfeeding</div>	<div>Rare infections during the first year of life</div> <div>High socioeconomic status</div> <div>Prenatal chromosomal translocations</div> <div>Agricultural occupation of parents</div>

Figure 12. Protective and risk factors for childhood acute lymphoblastic leukemia.

Rare infections during the first year of life are frequently reported as a risk factor for childhood leukemias.^[21] Conversely, multiple infections during this period emerge as a protective factor. These observations are underlined by correlative data: a high socioeconomic state represents a risk factor, whereas crowded household conditions and many siblings emerge as protective factors. Cattle farming has been reported as an additional risk factor, whereas more than six months of breast feeding seem to reduce the risk.

Two additional sets of data deserve discussion: the frequent occurrence of specific chromosomal translocations in leukemic cells, often observed already prenatally.^[22] The

same types of chromosomal alterations have also been found in healthy individuals, although here their frequency appears to be very low. Another striking observation originates from the description of occasional small clusters of leukemic cases, specifically in regions where an influx of urban populations occurred in previously rural areas.^[23]

4.3.2.2. Possible Explanations

Three main hypotheses have been published to explain the epidemiological findings: Greaves^[24] speculated that there exists an insufficient maturation state of the immune system in the case of low exposure to infections. Preceding chromosomal translocations as the first event, followed by delayed infection “with an unspecified agent” should increase the risk for subsequent leukemic conversion. Alternatively, Kinlen^[25] proposed that sudden mixing of a population of low exposure to a putative leukemogenic agent (particularly in rural areas) with another population originating from urban areas previously highly exposed to the incriminated agent could promote an epidemic of the relevant infection. These hypotheses were supplemented by a further speculation: assuming that the protective effect of multiple infections during the first year of childhood were due to the reduction of the load of a putative leukemogenic agent by interferon production as outlined in Figure 13.^[21,26]

Reports on the supertransforming properties of specifically replication-incompetent SV40 and murine polyomaviruses,^[27,28] in addition to the recent demonstration of replication-incompetent Merkel cell polyomavirus in Merkel cell carcinomas,^[4] resulted in an attempt to combine the three hypotheses, assuming that replication-incompetent polyoma-type viruses and high multiplicities at the time of initial infection represent an important precondition for an increased leukemogenic risk. The generation of replication-incompetent viral progeny seems to depend on high multiplicities of infection and the co-infection of cells with both replication-competent and incompetent genomes. The sole subsequent infection of a susceptible cell with a replication-

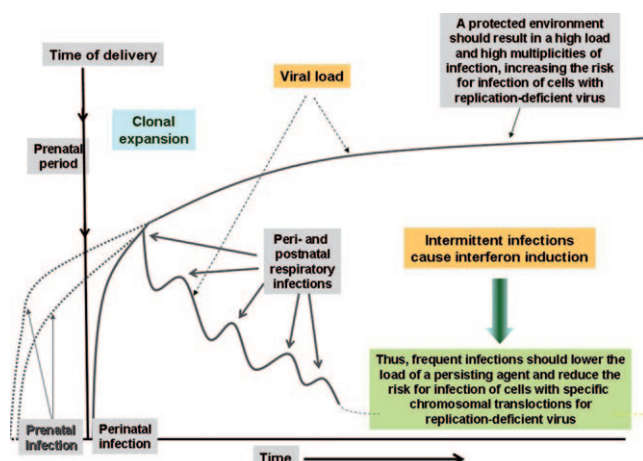


Figure 13. Schematic outline of the target cell conditioning hypothesis. Interferon synthesis resulting from multiple infections in early childhood reduces the load of a persisting potentially leukemogenic agent and thus reduces the risk of malignant proliferation.^[26]

incompetent genome may lead to the outgrowth of a leukemic clone. Susceptibility of a cell for this malignant conversion would require the previous or subsequent acquisition of a specific chromosomal translocation. These translocations also occur in healthy individuals, although at low frequency.^[29–32] They represent risk factors, but are clearly not sufficient for cell transformation. They should activate the oncogene of the replication-incompetent virus. A synopsis of this hypothesis is presented in Figure 14.

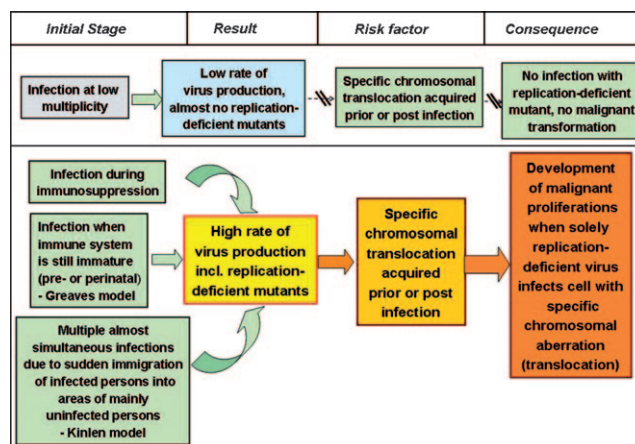


Figure 14. Synopsis of the target cell conditioning model for childhood leukemia.

A polyoma-type virus infection would fit best for this model, although members of structurally related virus families might also be considered. Since a number of reports document elevated risks in families of cattle farmers and for individuals in close contacts with cattle,^[21] at least part of childhood leukemias could be due to a native cattle virus. This virus should be replication-incompetent for human cells, but its oncogene may become activated in cells with specific chromosomal modifications. Since a number of reports also suggest human occupational risks of persons with communicative contacts (for example, teachers, hairdressers), other types of similar infections may be spread by human–human contacts.^[21]

It remains an interesting question to what extent other hematopoietic malignancies, like acute and chronic myelogenous leukemias, chronic lymphatic leukemias, B- and T-cell non-Hodgkin lymphomas, Epstein–Barr virus-negative Hodgkin lymphomas, and multiple myelomas could be included in these considerations. As yet undefined polyoma-virus-like particles have been electron microscopically demonstrated in trichodysplasia of a patient with non-Hodgkin's lymphoma.^[32]

5. Cancers Potentially Linked to Animal–Human Transmission

5.1. Colorectal, Breast, and Lung Cancers

A large number of reports consistently describe an increased risk for colorectal cancers related to a high

consumption of red meat.^[34,35] Recently this has also been noted for lung cancer in nonsmokers,^[36–38] and, to a more limited degree less consistently, also for breast cancer.^[37,39–42] A correlation seems to exist between countries with a high rate of red meat consumption and a high risk of colorectal and breast cancer. Common and frequently cited interpretations of these observations are dietary factors. Carcinogenic *N*-nitroso compounds, heterocyclic amines, and heterocyclic aromatic hydrocarbons arise during cooking, broiling, or meat curing. Some of these compounds require metabolic activation prior to converting into a carcinogenic form, as initially described by Sugimura and colleagues.^[43] In addition, potentially carcinogenic nitrosyl haem and nitroso thiols have been reported to be significantly increased in feces following a diet rich in red meat.^[44]

In contrast to red meat, consumption of white meat, and here specifically chicken and other poultry meat, has not been found to be associated with an elevated risk for colorectal or other cancers. It has been reported, however, that fried, grilled, or smoked chicken meat contains equally high concentrations of heterocyclic aromatic hydrocarbons and other carcinogens that arise in the preparatory steps prior to consumption.^[45–47] If this holds up and if no other hitherto unknown carcinogens are found specifically in red meat, these observations may require a fresh look at previous interpretations. In meat prepared medium or rare (Figure 15), temperatures in the central portions do not exceed 55 to 65 °C. At least some members of the polyoma- and papillomavirus families readily survive these temperatures without significant loss of their infectivity.^[48,49] The only known bovine polyomavirus was initially identified as a contamination of fetal bovine sera; thus, it must have been present in the peripheral blood of yet unborn or newborn calves. Existing members of the polyomavirus family have been poorly studied in our domestic animals. These viruses are commonly non-oncogenic in their natural hosts, but reveal carcinogenicity only in heterologous tissues. Currently six different genotypes of polyomaviruses have been identified in humans, but only one in cattle.

It is tempting to speculate that a hitherto unidentified bovine infectious agent with pronounced thermostability,



Figure 15. Red meat cooked “rare”.

replication-incompetent for human cells, and possibly structurally related to the polyomavirus family may play a role in colorectal cancer, potentially also in lung cancers of non-smokers and in breast cancer. This could be interpreted to mean that the described chemical carcinogens arising during cooking or curing processes are not sufficient for the induction of the respective cancers. In the case of red meat consumption they may, however, interact with viral agents, present in red, but not in white, meat.

6. Conclusions

Although we know that currently slightly more than 20 % of the global cancer incidence is linked to infectious events, some epidemiological observations suggest that this percentage will increase in the future. The recognition that no cancer linked to infections develops without additional modifications within the host-cell genome permits the speculation that even cancers with well-established chromosomal modifications deserve careful analysis for an additional involvement of infectious agents. Prime malignancies suggested here as candidates for potential links with infections are hematopoietic malignancies, particularly childhood lymphoblastic leukemias, Epstein–Barr virus-negative Hodgkin’s lymphomas, basal cell carcinomas of the skin, and breast, colorectal, and a subgroup of lung cancers. Although still hypothetical, this proposal is accessible to experimental verification. Even if only one of these speculations turns out to be correct, this would have profound implications for the prevention, diagnosis, and hopefully also for the therapy of the respective malignancy.

Received: March 19, 2009

Published online: July 8, 2009

- [1] a) H. zur Hausen, *Infections Causing Human Cancers*, Wiley-VCH, Weinheim, **2006**; b) “Global Cancer Statistics 2002”: D. M. Parkin, F. Bray, J. Ferlay, P. Pisani, *Ca-Cancer J. Clin.* **2005**, *55*, 74–108.
- [2] “Clonal integration of a polyomavirus in human Merkel cell carcinoma”: H. Feng, M. Shuda, Y. Chang, P. S. Moore, *Science* **2008**, *319*, 1096–1100.
- [3] “A children’s cancer dependent on climatic factors”: D. Burkitt, *Nature* **1962**, *194*, 232–234.
- [4] “T antigen mutations are a human tumor-specific signature for Merkel cell polyomavirus”: M. Shuda, H. Feng, H. J. Kwun, S. T. Rosen, O. Gjoerup, P. S. Moore, Y. Chang, *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 16272–16277.
- [5] “Cancer incidence before and after kidney transplantation”: C. M. Vajdic, S. P. McDonald, M. R. McCredie, M. T. van Leeuwen, J. H. Stewart, M. Law, J. R. Chapman, A. C. Webster, J. M. Kaldor, A. E. Grulich, *JAMA J. Am. Med. Assoc.* **2006**, *296*, 2823–2831.
- [6] “Human endogenous retrovirus family HERV-K(HML-2) RNA transcripts are selectively packaged into retroviral particles produced by the human germ cell tumor line Tera-1 and originate mainly from a provirus on chromosome 22q11.21”: K. Ruprecht, H. Ferreira, A. Flockerzi, S. Wahl, M. Sauter, J. Mayer, N. Mueller-Lantzsch, *J. Virol.* **2008**, *82*, 10008–10016.

- [7] "Identification of an infectious progenitor for the multiple-copy HERV-K human endogenous retroelements": M. Dewannieux, F. Harper, A. Richaud, C. Letzelter, D. Ribet, G. Pierron, T. Heidmann, *Genome Res.* **2006**, *16*, 1548–1556.
- [8] "Reconstitution of an infectious human endogenous retrovirus": Y. N. Lee, P. D. Bieniasz, *PLoS Pathog.* **2007**, *3*, e10.
- [9] "The replicative activity of human endogenous retrovirus K102 (HERV-K102) with HIV viremia": M. P. Laderoute, A. Giulivi, L. Larocque, D. Bellfo, Y. Hou, H.-X. Wu, K. Fowke, J. Wu, F. Diaz-Mitoma, *AIDS*, **2007**, *21*, 2417–2424.
- [10] "Epstein-Barr virus latent membrane protein LMP-2 A is sufficient for transactivation of the human endogenous retrovirus HERV-K18 superantigen": N. Sutkowski, G. Chen, G. Calderon, B. T. Huber, *J. Virol.* **2004**, *78*, 7852–7860.
- [11] "Negative thymocyte selection to HERV-K18 superantigens in humans": F. Meylan, M. De Smedt, G. Leclercq, J. Plum, O. Leupin, S. Marguerat, B. Conrad, *Blood* **2005**, *105*, 4377–4382.
- [12] "Cutting edge: Epstein-Barr virus transactivates the HERV-K18 superantigen by docking to the human complement receptor 2 (CD21) on primary B cells": F. C. Hsiao, M. Lin, A. Tai, G. Chen, B. T. Huber, *J. Immunol.* **2006**, *177*, 2056–2060.
- [13] H. zur Hausen, unpublished results.
- [14] "Human endogenous retrovirus K triggers an antigen-specific immune response in breast cancer patients": F. Wang-Johanning, L. Radvanyi, K. Rycak, J. B. Plummer, P. Yan, K. J. Sastry, C. J. Piyathilake, K. K. Hunt, G. L. Johanning, *Cancer Res.* **2008**, *68*, 5869–5877.
- [15] "Human endogenous retrovirus K (HML-2) elements in the plasma of people with lymphoma and breast cancer": R. Contreras-Galindo, M. H. Kaplan, P. Leissner, T. Verjat, I. Ferlenghi, F. Bagnoli, F. Giusti, M. H. Dosik, D. F. Hayes, S. D. Gitlin, D. M. Markovitz, *J. Virol.* **2008**, *82*, 9329–9336.
- [16] "Smallpox vaccination site complications": K. H. Waibel, D. S. Walsh, *Int. J. Dermatol.* **2006**, *45*, 684–688.
- [17] "Vaccinia virus, herpes simplex virus, and carcinogens induce DNA amplification in a human cell line and support replication of a helpervirus dependent parvovirus": J. R. Schlehofer, M. Ehrbar, H. zur Hausen, *Virology* **1986**, *152*, 110–117.
- [18] "Amplification of bovine papillomavirus DNA by *N*-methyl-*N*-nitro-*N*-nitrosoguanidine, ultraviolet irradiation, or infection with herpes simplex virus": J. Schmitt, J. R. Schlehofer, K. Mergener, L. Gissmann, H. zur Hausen, *Virology* **1989**, *172*, 73–81.
- [19] "Brain tumors in owl monkeys inoculated with a human polyomavirus (JC virus)": W. T. London, S. A. Houff, D. L. Madden, D. A. Fuccillo, M. Gravell, W. C. Wallen, A. E. Palmer, J. L. Sever, B. L. Padgett, D. L. Walker, G. M. ZuRhein, T. Ohashi, *Science* **1978**, *201*, 1246–1249.
- [20] "Proliferation-inducing viruses in non-permissive systems as possible causes of human cancers": H. zur Hausen, *Lancet* **2001**, *357*, 381–384.
- [21] "Childhood leukemias and other hematopoietic malignancies: Interdependence between an infectious event and chromosomal modifications": H. zur Hausen, *Int. J. Cancer* **2009**, DOI: 10.1002/ijc.24365.
- [22] "Pre-natal origins of childhood leukemia": M. Greaves, *Rev. Clin. Exp. Hematol.* **2003**, *7*, 233–245.
- [23] "Childhood leukemia and population mixing": L. J. Kinlen, *Pediatrics* **2004**, *114*, 330–331.
- [24] "The causation of childhood leukemia: a paradox of progress?": M. Greaves, *Discovery Med.* **2006**, *6*, 24–28.
- [25] "Epidemiological evidence for an infective basis in childhood leukaemia": L. J. Kinlen, *Br. J. Cancer* **1995**, *71*, 1–5.
- [26] "Virus target cell conditioning model to explain some epidemiologic characteristics of childhood leukemias and lymphomas": H. zur Hausen, E. M. de Villiers, *Int. J. Cancer* **2005**, *115*, 1–5.
- [27] "Enhanced transformation of human fibroblasts by origin-defective simian virus 40": M. B. Small, Y. Gluzman, H. L. Ozer, *Nature* **1982**, *296*, 671–672.
- [28] "Site-directed mutagenesis of the polyomavirus genome: replication-defective large T mutants with increased immortalization potential": C. Roberge, M. Bastin, *Virology* **1988**, *162*, 144–150.
- [29] "BCL2 translocation frequency rises with age in humans": Y. Liu, A. M. Hernandez, D. Shibata, D. A. Cortopassi, *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 8910–8914.
- [30] "Occurrence of bcl-2 oncogene translocation with increased frequency in the peripheral blood of heavy smokers": D. A. Bell, Y. Liu, G. A. Cortopassi, *J. Natl. Cancer Inst.* **1995**, *87*, 223–224.
- [31] "Quantification of t(14;18) in the lymphocytes of healthy adult humans as a possible biomarker for environmental exposures to carcinogens": J. C. Fuscoe, R. W. Setzer, D. D. Collared, M. M. Moore, *Carcinogenesis* **1996**, *17*, 1013–1020.
- [32] "Long-term clonal persistence and evolution of t(14;18)-bearing B cells in healthy individuals": S. Roulland, P. Lebailly, Y. Lecluse, N. Heutte, B. Nadel, P. Gauduchon, *Leukemia* **2006**, *20*, 158–162.
- [33] "Viral-associated trichodysplasia in a patient with lymphoma: a case report and review": S. S. Osswald, K. B. Kulick, M. M. Tomaszewski, L. C. Sperling, *J. Cutaneous Pathol.* **2007**, *34*, 721–725.
- [34] "Dietary meat, endogenous nitrosation and colorectal cancer": G. G. Kuhnle, S. A. Bingham, *Biochem. Soc. Trans.* **2007**, *35*, 1355–1357.
- [35] "Processed meat and colorectal cancer: a review of epidemiologic and experimental evidence": R. L. Santarelli, F. Pierre, D. E. Corpet, *Nutr. Cancer* **2008**, *60*, 131–144.
- [36] "A prospective study of red and processed meat intake in relation to cancer risk": A. J. Cross, M. F. Leitzmann, M. H. Gail, A. R. Hollenbeck, A. Schatzkin, R. Sinha, *PLoS Med.* **2007**, *4*, e325.
- [37] "Canadian Cancer Registries Epidemiology Research Group. Meat and fish consumption and cancer in Canada": J. Hu, C. La Vecchia, M. DesMeules, E. Negri, L. Mery, *Nutr. Cancer* **2008**, *60*, 313–324.
- [38] "Intakes of red meat, processed meat, and meat mutagens increase lung cancer risk": T. K. Lam, A. J. Cross, D. Consonni, G. Randi, V. Bagnardi, P. A. Bertazzi, N. E. Caporaso, R. Sinha, A. F. Subar, M. T. Landi, *Cancer Res.* **2009**, *69*, 932–939.
- [39] "Meat consumption and risk of breast cancer in the UK Women's Cohort Study": E. F. Taylor, V. J. Burley, D. C. Greenwood, J. E. Cade, *Br. J. Cancer* **2007**, *96*, 1139–1146.
- [40] "Meat consumption, N-acetyl transferase 1 and 2 polymorphism and risk of breast cancer in Danish postmenopausal women": R. Egeberg, A. Olsen, H. Autrup, J. Christensen, C. Stripp, I. Tetens, K. Overvad, A. Tjønneland, *Eur. J. Cancer Prev.* **2008**, *17*, 39–47.
- [41] "Red meat consumption during adolescence among premenopausal women and risk of breast cancer": E. Linos, W. C. Willett, E. Cho, G. Colditz, L. A. Frazier, *Cancer Epidemiol. Biomarkers Prev.* **2008**, *17*, 2146–2151.
- [42] "Meat consumption, heterocyclic amines, NAT2, and the risk of breast cancer": L. I. Mignone, E. Giovannucci, P. A. Newcomb, L. Titus-Ernstoff, A. Trentham-Dietz, J. M. Hampton, E. J. Orav, W. C. Willett, K. M. Egan, *Nutr. Cancer* **2009**, *61*, 36–46.
- [43] "Carcinogenicity in mice and rats of heterocyclic amines in cooked foods": H. Ohgaki, H. Hasegawa, T. Kato, M. Suenaga, M. Ubukata, S. Sato, S. Takayama, T. Sugimura, *Environ. Health Perspect.* **1986**, *67*, 129–134.
- [44] "Red meat and colorectal cancer risk: the effect of dietary iron and haem on endogenous N-nitrosation": A. J. Cross, J. R. Pollock, S. A. Bingham, *IARC Sci. Publ.* **2002**, *156*, 205–206.

- [45] "Presence of nitrosable mutagen precursors in cooked meat and fish": M. Yano, K. Wakabayashi, T. Tahira, N. Arakawa, M. Nagao, T. Sugimura, *Mutat. Res.* **1988**, 202, 119–223.
- [46] "Analysis of 200 food items for benzo[a]pyrene and estimation of its intake in an epidemiologic study": N. Kazerouni, R. Sinha, C. H. Hsu, A. Greenberg, N. Rothman, *Food Chem. Toxicol.* **2001**, 39, 423–436.
- [47] "Polycyclic aromatic hydrocarbons (PAHs) in meat products and estimated PAH intake by children and the general population in Estonia": M. Reinik, T. Tamme, M. Roasto, K. Juhkam, T. Tenno, A. Kiis, *Food Addit. Contam.* **2007**, 24, 429–437.
- [48] "Inactivation of 12 viruses by heating steps applied during manufacture of a hepatitis B vaccine": P. N. Lelie, H. W. Reesink, C. J. Lucas, *J. Med. Virol.* **1987**, 23, 297–301.
- [49] "Testing thermal resistance of viruses": A. Sauerbrei, P. Wutzler, *Arch. Virol.* **2009**, 154, 115–119.

Improve Quality Save Energy Cut Costs



Save 19% with
continuation order!



The complete set Modern Drying Technologies
ISBN: 978-3527-31554-3, 2011



Vol. 1:
Computational Tools at Different Scales
ISBN: 978-3527-31556-7, 2007



Vol. 2:
Experimental Techniques
ISBN: 978-3527-31557-4, 2008



Vol. 3:
Product Quality and Formulation
ISBN: 978-3527-31558-1, 2009



Vol. 4:
Energy Savings
ISBN: 978-3527-31559-8, 2010



Vol. 5:
Process Intensification
ISBN: 978-3527-31560-4, 2011

41031803_gu

www.wiley-vch.de

Wiley-VCH Verlag GmbH & Co. KGaA · POB 10 11 61 · D-69451 Weinheim · Germany
Phone: 49 (0) 6201/606-400 · Fax: 49 (0) 6201/606-184 · E-Mail: service@wiley-vch.de



WILEY-VCH

